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Neonatal high pressure hydrocephalus is associated with elevation of pro-inflammatory cytokines IL-18 and IFN γ in cerebrospinal fluid

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Abstract

Background: In human neonatal high pressure hydrocephalus (HPHC), diffuse white matter injury and gliosis predispose to poor neuro-developmental outcome. The underlying mechanism for diffuse white matter damage in neonatal HPHC is still unclear. Analogous to inflammatory white matter damage after neonatal hypoxemia/ischemia, we hypothesized that pro-inflammatory cytokines could be involved in neonatal HPHC. If so, early anti-inflammatory therapy could ameliorate white matter damage in HPHC, before irreversible apoptosis has occurred. In HPHC and control neonates, we therefore aimed to compare cerebrospinal fluid (CSF) concentrations of IL18, IFNγ and sFasL (interleukin 18, interferon gamma and apoptosis marker soluble-Fas ligand, respectively).

Methods: In neonatal HPHC (n = 30) and controls (n = 15), we compared CSF concentrations of IL18, IFN γ and sFasL using sandwich ELISA. HPHC was grouped according to etiology: spina bifida aperta (n = 20), aqueduct stenosis (n = 4), and fetal intra-cerebral haemorrhage (n = 6). Neonatal control CSF was derived from otherwise healthy neonates (n = 15), who underwent lumbar puncture for exclusion of meningitis.

Results: In all three HPHC groups, CSF IL18 concentrations were significantly higher than control values, and the fetal intracranial haemorrhage group was significantly higher than SBA group. Similarly, in all HPHC groups CSF-IFN γ concentrations significantly exceeded the control group. In both HPHC and control neonates, CSF FasL concentrations remained within the range of reference values.

Conclusion: Independent of the pathogenesis, neonatal HPHC is associated with the activation of the pro-inflammatory cytokines (IL-18 and IFN γ) in the CSF, whereas CSF apoptosis biomarkers (sFasL) were unchanged. This suggests that anti-inflammatory treatment (in addition to shunting) could be helpful to preserve cerebral white matter.

Background

Since the introduction of innovative drainage valves and third ventricular endoscopy, neurosurgical treatment strategies for neonatal HPHC have improved. Nevertheless, HPHC is still associated with irreversible white matter damage and adverse neurological outcome [1-4]. After hypoxemia/ischemia, white matter damage consists of a diffuse, inflammatory pattern involving pro-inflammatory cytokines, oligodendrocytic injury, gliosis and myelin loss [5-7]. Pro-inflammatory cytokines are biologically active proteins produced by T cells, astrocytes and microglial cells. After cytokine release, immune cells invade the brain and subsequently activate astrocytes and microglial cells, which results in apoptosis and gliosis [5,8].

Especially, the immature central nervous system is vulnerable for inflammatory damage. This is attributed to the specific sensitivity of immature oligodendrocytes for microglial cells, glutamate and free radicals [9,10]. Although shunting will improve cerebral perfusion and prevent gliosis [11], shunting does not address inflammatory consequences. Thus, long-acting cytokines (released before shunting), could theoretically continue to damage oligodendrocytes after shunting [6,12]. In the neonatal CNS, inflammatory mechanisms may contribute to diffuse white matter damage not only in the periventricular regions, but also at a distance from the ventricles [9]. Hypoxemia/ischemia is associated with cytokine IL18 release and cystic white matter damage [6]. Cytokine IL18 can induce other pro-inflammatory cytokines, such as IFNy, IL-1 β and TNF α [13]. In contrast to elevated CSF IL18 concentrations which last for months, IL-1β and TNF α concentrations are only elevated for hours [14,15]. This may explain our previously reported negative association between CSF IL-1β concentration and cystic white matter damage [6]. Cytokine IFNy is also involved in the regulation of the inflammatory response by activation of cytotoxic T-cells and macrophages [16]. Upon activation, this may result in apoptosis, myelin loss and gliosis [5,12,17-20]. In neonates with post-hemorrhagic hydrocephalus and cystic white matter damage, we have subseshown that enhanced quently growth factor concentrations (i.e. vascular endothelial growth factor (VEGF) and transforming growth factor β 1 (TGF- β 1)) will finally reflect tissue repair [21,22].

In this perspective, we hypothesized that hypoxemia/ischemia-related up-regulation of longer acting cytokines (IL18 and IFNγ) could be involved in neonatal white matter damage by HPHC. If cytokines are involved in ongoing white matter damage, early anti-inflammatory therapy could be beneficial, before irreversible apoptosis has occurred. The apoptosis biomarkers Fas and Fas-ligand (FasL) are members of the tumour-necrosis factor super family. The death Fas (CD95/Apo-1) is located on the cell

surface. It plays a pivotal role in transduction of the apoptotic cell death program. Fas and its FasL exist in membrane bound form and soluble forms and can be detected in neonatal CSF [23]. Soluble FasL (sFasL) is expressed on activated T cells and released by metalloproteinase. SFasL can regulate extracellular apoptosis by pro- and anti-apoptotic properties. Expression of sFasL indicates ongoing apoptosis.

In neonatal HPHC characterized by progressive ventriculomegaly and increased head circumference > P_{75} , and control patients, this study aimed to determine and compare CSF IL18, IFN γ and sFasL concentrations. We hypothesized that CSF IL-18 and IFN γ concentrations are increased in HPHC, irrespective of underlying etiology. To investigate this, HPHC patients were grouped according to three different aetiologies: spina bifida aperta (SBA), aqueduct stenosis, and hydrocephalus after fetal intra-cranial haemorrhage.

Methods

Patients selection and CSF sampling

The study was approved by the medical ethical committees of the University of Bonn, the Charité Universitäts Medizin Berlin and the University Medical Center Groningen. After informed consent by the parents, 30 HPHC and 15 control neonates were included. In neonatal HPHC, CSF was obtained during initial neonatal shunt surgery. Indications for shunting consisted of clinical signs for high intracranial pressure, bulging fontanel, widening of the sagittal suture, progressive ventriculomegaly and increased head circumference (> P₇₅). Since anaesthesia, artificial respiration and internal pressure compensation may quantitatively influence the assessment of intracranial pressure, CSF pressure was not measured routinely during shunt placement. Neonatal HPHC was grouped according to aetiology: SBA (n = 20; characterized by presence of meningomyelocele), aqueduct stenosis (n = 4); HC after fetal intra-cranial haemorrhage (n = 6). Selection of HC after fetal intra-cranial haemorrhage (i.e. haemorrhage 4-6 weeks before delivery) allowed avoidance of the potentially confounding influence by disintegration of platelets. The diagnosis of fetal post-hemorrhagic hydrocephalus was confirmed by prenatal ultrasound (ATL 500, 3.5 MHz transducer), postnatal ultrasound (Vingmed Vivid5, multi-frequency transducer (5-7.5-10 MHz crystals) and magnetic resonance imaging (Philips Healthcare, Best, Netherlands, 1.5 Tesla). Low risk neonates, undergoing lumbar puncture for exclusion of meningitis, served as controls (n = 15). Gestational ages in the three hydrocephalic groups and control group were similar, i.e. between 27-54 and 24-54 weeks, respectively. CSF samples obtained during shunt revisions, neonatal asphyxia and CNS infections were excluded. Cerebral infection was excluded by negative CSF cultures,

cellular count, total protein concentration and by assessment of CSF-IL6 concentrations (in CNS infection, CSF IL-6 concentrations are increased). CSF-IL6 concentrations were measured by commercially available solid-phase enzyme-labelled chemiluminescent sequential immunometric assay on an Immulite analyzer (DPC Biermann, Bad Naunheim, Germany). All CSF IL-6 concentrations were within the normal range 5 pg/ml – 200 pg/ml (i.e. far below CSF IL-6 levels in newborns with bacterial ventriculitis [24]). Total CSF protein content varied between 0.1 – 2.5 g/l.

CSF analysis

All CSF samples were immediately centrifuged and stored at -40°C for further analysis. CSF concentrations of IL-18 and IFNγ were determined by sandwich ELISA (R&D systems, Wiesbaden, Germany) according to the manufacturer's instructions. The sensitivity of the assay was 12.5 pg/ml for IL18, 8.0 pg/ml for IFNγ and 0.5 ng/ml for sFasL using Mab for coating and binding (clones 4H9 and 4A5) [6,23]. The intra-assay coefficient of variation was 5.0% for IL18 and 4.7% for IFNγ. All ELISA 96-well micro titer plates were analyzed using a microplate photometer (Dynotech MR5000, Denkendorf, Germany). Neonatal control CSF sFasL data were derived from our previous study by application of the same analytical technique, performed by the same laboratory [23].

Data analysis

For statistical analysis, Mann-Whitney U test was used with two-sided p values to compare continuous nonparametric group of values, as the distribution of values was non-Gaussian.

Results

Irrespective of the underlying cause, IL-18 concentrations were significantly higher in HPHC neonates than in controls, median and range: SBA: 80 (23-232) pg/ml; aqueduct stenosis: 66 (55-226) pg/ml; fetal intracranial haemorrhage: 223 (103-406) pg/ml; controls: 12.5 (12.5-158) pg/ml. Each group was significantly higher than control, p < 0.01, and the fetal intracranial haemorrhage group was significantly higher than SBA, p < 0.01; figure 1A). Similarly, CSF IFNy concentrations were also significantly higher in the three HPHC groups than in controls, median and range: SBA: 35 (12-139) pg/ml; aqueduct stenosis: 22 (15-28) pg/mL; fetal intracranial haemorrhage: 22 (17-56) pg/mL; controls: 8 (8-22) pg/ ml. Each group was significantly higher than controls, p <0.01; but not significantly different between the groups (figure 1B). In all three neonatal HPHC groups, CSF sFasL concentrations remained within control limits, < 0.5 ng/ ml [23].

Discussion

Under diverse cerebral pathological circumstances, both astroglial and microglial alterations may be involved in white matter damage and adverse neurological outcome. It is indicated that hydrocephalus-associated brain tissue compression can instigate proliferation of astrocytes and microglial cells resulting in gliosis [11]. This study has shown that irrespective of the underlying aetiology, early indications for HPHC (derived from concurring ventriculomegaly and macrocephaly) are accompanied by proinflammatory cytokine activation (IL-18 and IFNy) with highest IL18 concentrations in post-hemorrhagic HPHC. Despite cytokine release into the CSF, the CSF concentrations of the apoptosis biomarker sFasL remained within control limits. These results are contrasted by our previous findings of high CSF sFasL concentrations in neonatal cystic white matter damage [23]. In the present study, normal CSF sFasL concentrations are explained by early assessment of CSF samples during the first shunt implantation and before cystic white matter alterations have occurred. All together in early neonatal HPHC, present data indicate that inflammation precedes irreversible apoptosis, which may provide a theoretical basis for early anti-inflammatory therapy (at about the time of first shunt implantation). In children with leucomalacia and post-haemorrhagic hydrocephalus, similar cytokine activation is associated with a diffuse component of white matter damage, prolonged myelination delay (for months) and even permanent myelin deficiency [5,6]. From a neuro-pathological point of view, concurrent white matter lesions of varying appearance and age (acute, organizing and chronic) suggest various, ongoing insults in the same patient [5]. However, before these data can be extrapolated to all groups of neonatal HPHC, histological examination (by immunostaining) will be required. Analogous to pediatric HPHC, adult patients with normal pressure hydrocephalus and/or vascular dementia may also have elevated pro-inflammatory cytokine concentrations (TNFα) in association with white matter damage [25,26]. However, because of patient heterogeneity, agespecific cytokine sensitivity and variability in disease progression, it is not possible to speculate further about similarities in inflammatory involvement between ages.

In neonatal H-Tx rat (i.e. an animal model for congenital hydrocephalus by aqueduct stenosis), it was shown that shunting could ameliorate gliosis [11]. Since gliosis may be associated with both reactive astrocytosis and microgliosis, one would expect that anti-inflammatory therapy could have a beneficial effect in addition to shunting. Accordingly, it was shown that minocycline, a semi-synthetic second generation tetracycline with anti-inflammatory, anti-apoptotic and anti-glutaminergic properties

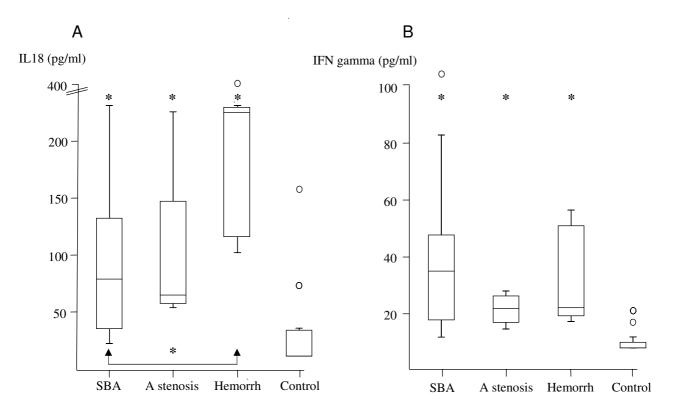


Figure I (A) Graphs of CSF interleukin-18 (IL-18) concentration and (B) CSF interferon gamma (IFN gamma) concentration in CSF from neonatal HPHC. The vertical axes indicate concentration (pg/ml). The horizontal axes indicates three different age-matched aetiologies for neonatal HPHC: spina bifida aperta (SBA), aqueduct stenosis (A stenosis), and fetal intracranial hemorrhage (Hemorrh) and neonatal controls (Control). Data are median and range plus 25^{th} and 75^{th} percentiles Encircled symbols in the figures indicate single parameters that appeared out of range. A: In all three neonatal HPHC groups, IL-18 concentrations were significantly higher than in controls (* p < 0.01). Furthermore, the fetal intracranial hemorrhage hydrocephalus group was significantly higher than the SBA hydrocephalus group (indicated by arrows at the bottom * p < 0.01). B: In all three neonatal HPHC groups, CSF IFN γ concentrations were significantly higher than in controls (* p < 0.01).

[27], reduces gliotic scarring in H-Tx rat [28]. Although minocycline is contra-indicated in young children, present human neonatal HPHC data suggest that other anti-inflammatory compounds could theoretically ameliorate diffuse cytokine-coupled, white matter damage [11]. In multiple sclerosis (characterized by up-regulation of pro-inflammatory cytokines), different anti-inflammatory agents (such as interferon beta (IFNβ) and glatiramer acetate) are known to ameliorate white matter damage [29]. Although there may be a rational basis for early neonatal (or perhaps even fetal) application of such anti-inflammatory compounds, potentially harmful adverse reactions should first be considered.

Conclusion

Neonatal HPHC irrespective of cause, is accompanied by pro-inflammatory cytokine activation (IL-18 and IFN γ) in the CSF. These data suggest that anti-inflammatory treat-

ment (in addition to shunting) could be helpful to preserve cerebral white matter in these patients.

Abbreviations

CSF: cerebrospinal fluid; GA: gestational age; HPHC: neonatal high pressure hydrocephalus; IFN β : interferon beta; IFN γ : interferon gamma; IL18: interleukin 18; sFas: soluble Fas ligand; SBA: spina bifida aperta.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

DS contributed to the study design, collection of CSF and writing of the manuscript. UF contributed to the study design, assessment of CSF samples and correction of the manuscript. TS contributed to the assessment of CSF samples and correction of the manuscript. EH contributed to

collection of CSF and correction of the manuscript. CS contributed to collection of CSF and correction of the manuscript. AH contributed to the study design, collection of CSF, statistical analysis, and helped drafting and correcting the manuscript. All authors have read and approved the final version of the manuscript.

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References

- Johanson CE, Duncan JA III, Klinge PM, Brinker T, Stopa EG, Silverberg GD: Multiplicity of cerebrospinal fluid functions: New challenges in health and disease. Cerebrospinal Fluid Res 2008, 5:10.
- Del Bigio MR: Pathophysiologic consequences of hydrocephalus. Neurosurg Clin N Am 2001, 12:639-49. vii
- Del Bigio MR: Future directions for therapy of childhood hydrocephalus: a view from the laboratory. Pediatr Neurosurg 2001. 34:172-181.
- Del Bigio MR: Neuropathological changes caused by hydrocephalus. Acta Neuropathol (Berl) 1993, 85:573-585.
- Folkerth RD: Neuropathologic substrate of cerebral palsy. J Child Neurol 2005, 20:940-949.
- Schmitz T, Heep A, Groenendaal F, Huseman D, Kie S, Bartmann P, Obladen M, Felderhoff-Muser U: Interleukin-Ibeta, interleukin-18, and interferon-gamma expression in the cerebrospinal fluid of premature infants with posthemorrhagic hydrocephalus – markers of white matter damage? Pediatr Res 2007, 61:722-726.
- Cherian S, Whitelaw A, Thoresen M, Love S: The pathogenesis of neonatal post-hemorrhagic hydrocephalus. Brain Pathol 2004, 14:305-311
- Folkerth RD, Keefe RJ, Haynes RL, Trachtenberg FL, Volpe JJ, Kinney HC: Interferon-gamma expression in periventricular leukomalacia in the human brain. Brain Pathol 2004, 14:265-274.
- Folkerth RD: Periventricular leukomalacia: overview and recent findings. Pediatr Dev Pathol 2006, 9:3-13.
- Volpe JJ: Neurobiology of periventricular leukomalacia in the premature infant. Pediatr Res 2001, 50:553-562.
- Miller JM, McAllister JP: Reduction of astrogliosis and microgliosis by cerebrospinal fluid shunting in experimental hydrocephalus. Cerebrospinal Fluid Res 2007, 4:5.
- Town T, Nikolic V, Tan J: The microglial "activation" continuum: from innate to adaptive responses. J Neuroinflammation 2005. 2:24.
- Okamura H, Tsutsi H, Komatsu T, Yutsudo M, Hakura A, Tanimoto T, Torigoe K, Okura T, Nukada Y, Hattori K: Cloning of a new cytokine that induces IFN-gamma production by T cells. Nature 1995, 378:88-91.
- Szaflarski J, Burtrum D, Silverstein FS: Cerebral hypoxia-ischemia stimulates cytokine gene expression in perinatal rats. Stroke 1995, 26:1093-1100.
- Hedtjarn M, Leverin AL, Eriksson K, Blomgren K, Mallard C, Hagberg H: Interleukin-18 involvement in hypoxic-ischemic brain injury. J Neurosci 2002, 22:5910-5919.
- Degliantoni G, Murphy M, Kobayashi M, Francis MK, Perussia B, Trinchieri G: Natural killer (NK) cell-derived hematopoietic colony-inhibiting activity and NK cytotoxic factor. Relationship with tumor necrosis factor and synergism with immune interferon. J Exp Med 1985, 162:1512-1530.
- Baerwald KD, Popko B: Developing and mature oligodendrocytes respond differently to the immune cytokine interferon-gamma. J Neurosci Res 1998, 52:230-239.
- Mana P, Linares D, Fordham S, Staykova M, Willenborg D: Deleterious role of IFNgamma in a toxic model of central nervous system demyelination. Am J Pathol 2006, 168:1464-1473.
- Hirsch RL, Panitch HS, Johnson KP: Lymphocytes from multiple sclerosis patients produce elevated levels of gamma interferon in vitro. J Clin Immunol 1985, 5:386-389.

- Corbin JG, Kelly D, Rath EM, Baerwald KD, Suzuki K, Popko B: Targeted CNS expression of interferon-gamma in transgenic mice leads to hypomyelination, reactive gliosis, and abnormal cerebellar development. Mol Cell Neurosci 1996, 7:354-370.
- Heep A, Bartmann P, Stoffel-Wagner B, Bos A, Hoving E, Brouwer O, Teelken A, Schaller C, Sival D: Cerebrospinal fluid obstruction and malabsorption in human neonatal hydrocephaly. Childs Nerv Syst 2006, 22:1249-1255.
- Heep A, Stoffel-Wagner B, Bartmann P, Benseler S, Schaller C, Groneck P, Obladen M, Felderhoff-Mueser U: Vascular endothelial growth factor and transforming growth factor-betal are highly expressed in the cerebrospinal fluid of premature infants with posthemorrhagic hydrocephalus. Pediatr Res 2004, 56:768-774.
- Felderhoff-Mueser U, Buhrer C, Groneck P, Obladen M, Bartmann P, Heep A: Soluble Fas (CD95/Apo-1), soluble Fas ligand, and activated caspase 3 in the cerebrospinal fluid of infants with posthemorrhagic and nonhemorrhagic hydrocephalus. Pediatr Res 2003, 54:659-664.
- Baumeister FA, Pohl-Koppe A, Hofer M, Kim JO, Weiss M: IL-6 in CSF during ventriculitis in preterm infants with posthemorrhagic hydrocephalus. *Infection* 2000, 28:234-236.
- Tarkowski E, Tullberg M, Fredman P, Wikkelso C: Normal pressure hydrocephalus triggers intrathecal production of TNF-alpha. Neurobiol Aging 2003, 24:707-714.
- Tarkowski E, Tullberg M, Fredman P, Wikkelso C: Correlation between intrathecal sulfatide and TNF-alpha levels in patients with vascular dementia. Dement Geriatr Cogn Disord 2003. 15:207-211.
- 27. Maier K, Merkler D, Gerber J, Taheri N, Kuhnert AV, Williams SK, Neusch C, Bahr M, Diem R: Multiple neuroprotective mechanisms of minocycline in autoimmune CNS inflammation. Neurobiol Dis 2007, 25:514-525.
- Miller JM, Shanku AG, Ham SD, McAllister JP: Inhibitory effects of minocycline on gliosis in the hydrocephalic H-Tx rat [abstract]. Cerebrospinal Fluid Res 2006, 3:s17.
- Živadinov R, Reder AT, Filippi M, Minagar A, Stuve O, Lassmann H, Racke MK, Dwyer MG, Frohman EM, Khan O: Mechanisms of action of disease-modifying agents and brain volume changes in multiple sclerosis. Neurology 2008, 71:136-144.

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